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Title:	LANL IBC Meeting Minutes, June 5, 2025
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LANL IBC Meeting Minutes, June 5, 2025		
Element		**Notes**
Institution	*Los Alamos National Laboratory*	
Meeting Date	*Thursday, June 5, 2025*	
Meeting Time	*8:33 AM – 10:36 AM*	
Meeting Type	*In person meeting and Virtual via Webex*	
IBC Members	1. Kumkum Ganguly, (IBC Chair/ Biology) 2. Georgia Ali, (BSO/ Industrial Hygiene/Biology) 3. Carla Jo Logan Young, (BSO back-up/ Industrial Hygiene/Biology) 4. Sara Pasqualoni, (SOMD) 5. Armand Dichosa, (IBC Member/ Biology) 6. Sofiya N. Micheva-Viteva, (IBC Member/ Biology) 7. Jessica Cubicek Sutherland (IBC Member/Biology) 8. Maureen Dolan, (Non-voting Member/Observer - Legal) 9. Wesley David Boose, MD (IBC Member/ Occupational Medicine) 10. Kent Allen Candee, (IBC Member/ Industrial Hygiene) 11. Richard Honsinger, MD (Non-affiliated Community Member; Los Alamos Medical Center) 12. Joyce Ritchins, RN (Non-affiliated Community Member; Los Alamos Medical Center) 13. Tamas Torok, (Other DOE Member; Lawrence Berkeley National Laboratory)	
Quorum	*The IBC has 12 voting members, and 1 non-voting member. For a quorum, 8 members are required to conduct business. Late arrivals and early departures to be noted here.*	**Quorum present.** Jessica Z. Kubicek-Sutherland, Maureen Dolan, and Sara Pasqualoni were absent and Sofiya N. Micheva-Viteva left early.
Other Individuals in Attendance		None
Call to Order		**IBC Action:** Call to order 8:33 AM
Review and approval of previous meeting minutes		**IBC Action:** March 6th, 2025 meeting minutes approval Votes: 10 members - For · For/Against/Abstain: (9 For/ 0 Against/ 0 Abstain)
Review of Prior Business		· Discussion of meeting minutes, including changes related to Robert's Rules of Order · Plan to use the NIH Meeting Minutes Template, starting September 2025
New IBC Registration for Review		
PI Name(s)	*Name (s) Kirti Bharjwaj, Apoorv Shanker, Maria Nevarez Martinez*	
Registration Number/Title	*New Registration 2025: IBC-198* *Auger Emitters for Cancer Treatement*	Making Safer Radiation Accessible for Cancer Treatments
Project Overview	• **Agent name:** Human cancer cell lines; BT-474, MCF7 (Two cell types — BT-474 cells and MCF-7 cells are breast cancer cells with epithelial morphology – will be used following standardized protocols) • **Agent Characteristics:** BT-474 cells are breast cancer cells with epithelial morphology. MCF-7 cells are breast cancer cells with epithelial morphology. • **Sources and nature of the nucleic acid sequences (e.g., species, structural transgene, oncogene, toxin):** None. • **Host(s), vector(s), and Donor Genes if used:** None • **Modifications (e.g., deletions, insertions, mutations to attenuate, or render replication incompetent) and note of any supporting documentation (published or unpublished data):** None • **Types of experimental manipulations that will be employed:** cell culturing and use of radioactive labeled nanoparticles. • **Proposed biosafety containment levels at which each operation will occur:** BSL-2	• **Additional pertinent information:** · The PI is proposing to develop and validate nano-based chealators to effectively deliver Auger electron emitting radionuclides to tumor cells for targeted auger electron therapy. · All cell line work will be manipulated under the BSL-2 conditions inside a Class II Biosafety Cabinet with use of appropriate PPE and methods. · Section III-D-1-a - the protocol uses commercial cell lines have been tested for well-known bloodborne pathogens, including HIV-1, HCV and HBV. Human cell lines could be potential carriers of poorly identified cancer-inducing viral pathogens. Therefore, these cells shall be handled as containing potentially infectious agents using universal precautions. All human cell lines are considered BSL-2 at LANL and are subject to the provisions of the BBP Standard.
NIH Guidelines Section	**III-D-1-a: Experiments involving the introduction of recombinant or synthetic nucleic acid molecules into Risk Group 2 agents will usually be conducted at Biosafety Level (BL) 2 containment.**	Two cell types — BT-474 cells and MCF-7 cells are breast cancer cells with epithelial morphology – will be used following standardized protocols
Risk Assessment and Discussion	• Individuals will wear Standard PPE for BSL-2 labs: a disposable long-sleeved lab coat, safety glasses, and nitrile gloves. Two pairs of gloves (typically one layer of nitrile/neoprene and one layer of vinyl) are worn and disposable sleeves. Hand washing is required before leaving the BSL-2 lab. Staff will be trained in laboratory safety practices, including sharps safety. • No sharps will be used in the BSL-2 labs for any of the activities. • PPE for Rad: The outer/second layer of gloves, typically made of PVC will be disposed of in the rad trash after use. When entering the biosafety cabinet, disposable Tyvek sleeve will be used. Also, a personal dosimetry device (TLD) will be used while working. • Cells will be cultured in disposable sterile flasks using disposable filtered pipettes for all manipulations to protect against aerosols.	Pathogenicity: All commercial cell lines have been tested for well-known bloodborne pathogens, including HIV-1, HCV and HBV. Though, human cell lines could be potential carriers of blood bourne pathogens. All human cell lines are considered BSL-2 at LANL and are subject to the provisions of the BBP Standard. Route of transmission: Bloodborne or inhalation of aerosols. Environmental stability: Cells can be rendered non-viable using 2-4% paraformaldehyde for 15min, or 70% ethanol. Cells can be lysed using 1% detergent solution, and do not survive longer than 3 h without 5% CO2 supplementation at 37 °C. Cell lines are stable when frozen in LN2 in complete growth media with 5% DMSO. Infectious dose: Concentration/Total volume: To be purchased from ATCC. Frozen stocks will be stored at 106 cells/ml in a volume of 1 mL per aliquot (<10 aliquots in total). Availability of effective medical preventative or treatment options: Exposure shall be reported immediately to Occupational Medicine and therapeuticswill be determined by physician.
Training	**Document completion of required institutional level training as well as detailed laboratory or protocol specific training.**	· Current LANL Biosafety Training · Current LANL BSL2 Profiency checklist on file · Current LANL Chemical Worker · Current LANL Rad-Worker II Training · Safe sharps handling
Occupational Health Representative review (if applicable):	**Note any:** • Vaccination requirements • Respiratory protection • Periodic review of any medical surveillance • Post-exposure response procedures	Bloodborne Pathogen Medical Surveillance Enrollment, Human Pathogen Medical Surveillance Enrollment, Exposure shall be reported immediately to Occupational Medicine and prophylaxis will be determined by physician.
Biosafety Level Assignment	• BSL2 labs for all cell line work will be manipulated inside a Class II Biosafety Cabinet with use of appropriate PPE and methods.	· Individuals will wear Standard PPE for BSL-2 labs: a disposable long-sleeved lab coat, safety glasses, and nitrile gloves. Two pairs of gloves (typically one layer of nitrile/neoprene and one layer of vinyl) are worn and disposable sleeves.
	• Mixed Waste Requirements	All waste materials removed from the biosafety cabinet will be disinfected with 10% bleach solution and solidified with waste lock before going into the radiological waste. These include used laboratory plasticware and gloves contaminated with mammalian cells, and culture media.
IBC Vote	***Note:*** *If the IBC grants approvals based on specific conditions being met, there should be a formal mechanism for verifying the conditions are fulfilled (e.g., the BSO will conduct an inspection to verify all Biological Safety Cabinets are up to date on certification before work may commence, all training must be completed before lab staff may begin work etc.).*	IBC Action: Votes: 10 members- For- For/Against/Abstain: (9 For/ 0 Against/ 0 Abstain) - Conflict(s) of Interest: None. - The motion to approve the registration pending the following changes or conditions to be met was made and was seconded.
IBC Registrations for Review: Renewal		
PI Name(s)	*Name (s) Emilia Solomon, Kumkum Ganguly, and Sofiya Micheva-Viteva*	
Registration Number/Title	*Renewal Registration 2025: IBC-145* *Neuromuscular Junction Research*	Unsupervised Tensor Factorization AI Platform for Discovery of Broad-Spectrum Antiviral Targets from Global Omics Data and Milagro Goat Farm project
Project Overview	• **Agent name:** NSC34, C2C12, HSKMC, HSMM, WA09, NSC-H9, H9-SynGFP, WTC11/GM25256, NHEK, NHDF. • **Agent Characteristics:** Immortilazed human cell lines and stem cells and primary cells. • **Sources and nature of the nucleic acid sequences (e.g., species, structural transgene, oncogene, toxin):** Cedarlane (CLU140), ATCC, Lifeline, Lonza, WiCell, WiCell, WiCell, Coriell Institute, (Lonza and ATCC), (Lonza and ATCC), respectively. • **Host(s), vector(s), and Donor Genes if used:** None • **Modifications (e.g., deletions, insertions, mutations to attenuate, or render replication incompetent) and note of any supporting documentation (published or unpublished data):** None • **Types of experimental manipulations that will be employed:** cell culturing and small molecule exposure. • **Proposed biosafety containment levels at which each operation will occur:** BSL-2	• **Additional pertinent information:** · The PI is proposing to investigate inflammatory responses of the blood-brain barrier and brain organoid models to treatments with small molecule compounds, developed as anti-viral therapeutics and to investigate changes in skin models treated with goat milk soap using mass spec analysis and standard cell biology assays. · All cell line work will be manipulated under the BSL-2 conditions inside a Class II Biosafety Cabinet with use of appropriate PPE and methods. This should protect any personnel who are working with these cell lines. · Section III-D-1-a - the protocol uses commercial cell lines have been tested for well-known bloodborne pathogens, including HIV-1, HCV and HBV. Human cell lines could be potential carriers of poorly identified cancer-inducing viral pathogens. Therefore, these cells shall be handled as containing potentially infectious agents using universal precautions. All human cell lines are considered BSL-2 at LANL and are subject to the provisions of the BBP Standard.
NIH Guidelines Section	**III-D-1-a: Experiments involving the introduction of recombinant or synthetic nucleic acid molecules into Risk Group 2 agents will usually be conducted at Biosafety Level (BL) 2 containment.**	Six cell types–brain microvascular endothelial cells (BMEC), neuronal crest-derived pericyte-like cells (ncPC), astrocytes, cortical neurons (CN) microglia and macroglia – will be developed from human embryonic stem cell line WA09 following standardized protocols
Risk Assessment and Discussion	• During the centrifugation process the aerosol-tight covers will be screwed on the rotor buckets. Standard sterile practices will be used during cell culture with a regular (weekly/monthly) equipment cleaning schedule. • Individuals will wear Standard PPE for BSL-2 labs: consisting of disposable lab coat, safety (add glasses) or goggles and nitrile gloves must be always donned in the BSL2 room and must be removed upon exiting the lab. Hand washing is required before leaving the BSL-2 room. Staff will be trained in laboratory safety practices, including sharps safety. • Cells will be cultured in disposable sterile flasks using disposable filtered pipettes for all manipulations to protect against aerosols.	Pathogenicity: All commercial cell lines have been tested for known blood-borne pathogens, including HIV-1, HCV and HBV. Human cell lines could be potential carriers of poorly identified cancer inducing viral pathogens. Therefore, the cell lines shall be handled as containing potentially infectious agents using universal precautions. Route of transmission: Direct contact of skin or mucous membranes, ingestion, and accidental parenteral inoculation are the primary laboratory hazards associated with cell cultures of immortalized cell lines or primary cells. The most likely route for personnel handling cell culture samples is dermal exposure. Environmental stability: Human and Animal cell lines can be killed by fixation for 30 min with 2% paraformaldehyde, 10% bleach, or 70% ethanol. They can be lysed with 1% of detergent and do not survive atmospheric O2 pressure for longer than 8h. Cell lines are stable frozen in LN2 in 10% serum and 10% DMSO, Infectious dose: Not Applicable, Availability of effective medical preventative or treatment options: Exposure is reported immediately to occupational medicine. Therapeutics will be determined by a
Training	**Document completion of required institutional level training as well as detailed laboratory or protocol specific training.**	· Current LANL Biosafety Training · Current LANL BSL2 Profiency checklist on file · Current LANL Chemical Worker: Trizol and Formaldehyde Proficiency · Current LANL Rad-Worker II Training · Safe sharps handling
Occupational Health Representative review (if applicable):	**Note any:** • Vaccination requirements • Respiratory protection • Periodic review of any medical surveillance • Post-exposure response procedures	Bloodborne Pathogen Medical Surveillance Enrollment, Human Pathogen Medical Surveillance Enrollment, Exposure shall be reported immediately to Occupational Medicine and prophylaxis will be determined by physician.
Biosafety Level Assignment	• BSL2 labs for all cell line work will be manipulated inside a Class II Biosafety Cabinet with use of appropriate PPE and methods.	· Individuals will wear Standard PPE for BSL-2 labs: a disposable long-sleeved lab coat, safety (add glasses) or goggles, and nitrile gloves. Two pairs of gloves (typically one layer of nitrile/neoprene and one layer of vinyl) are worn and disposable sleeves.
	• Standard Biological Waste Requirements	· Waste will be closed, and the outer surface will be decontaminated with 10% bleach before removal from the BSC. The decontaminated bag is then transferred to the autoclave in a secondary plastic container. Liquid waste will be inactivated by adding undiluted bleach for overnight incubation followed by dilution to a final bleach concentration of a 10% bleach solution (100mL bleach + 900mL liquid waste).
IBC Vote	*Note: If the IBC grants approvals based on specific conditions being met, there should be a formal mechanism for verifying the conditions are fulfilled (e.g., the BSO will conduct an inspection to verify all Biological Safety Cabinets are up to date on certification before work may commence, all training must be completed before lab staff may begin work etc.).*	IBC Action: Votes: 8 members - For; 1 member-Abstain- For/Against/Abstain: (8 For/ 0 Against/ 1 Abstain) - Conflict(s) of Interest: Committee Member Name(s): Kumkum Ganguly and Sofiya N. Micheva-Viteva - The motion to approve the registration pending the following changes or conditions to be met was made and was seconded.
New Business/ Additional Topics	The BSO and IBC chair received confirmation from NIH regarding the new format for the IBC Committee Meeting minutes that are effective June 2025.	The approved IBC meeting minutes will be posted in LANL public website.
Review of Incidents	*The NIH Guidelines require that significant incidents, violations and research-related accidents and illnesses be reported to NIH OSP.* *For information regarding incident reporting requirements please refer to the Incident Reporting FAQs.*	No Incidents were reported.
Inspections/ Ongoing Oversight	*For IBC -194 and IBC-197 met Inspections for the labs granted approval.*	
Public Comments	There were no public comments.	
Adjournment		IBC Action: Adjournment 10:36 AM Votes: 9 members - For- For/Against/Abstain: (9 For/ 0 Against/ 0 Abstain) - The next meeting scheduled is for September 4th, 2025 from 8:30 am to 11:30 am in person and via Teams/WebEx.